Review article on "Leprosy (Hansen's Disease)" and "Indian Medicinal Plants Used In treatment of Leprosy"

1st Sahil Nasirkhan Pathan, 2nd Pritam Deshmukh, 3rd Rohit Singh, 4th Shoaib Pathan, 5th Sneha Rodge

¹Student, ²Student, ³Student, ⁴Student, ⁵Student

¹Central India College of Pharmacy, Lonara, Nagpur 441111, Maharashtra, India

Abstract

Leprosy is a neglected infectious disease caused by the acid-fast bacterium Mycobacterium leprae. A deeper understanding of the structural and biological features of M. leprae and the genomic sequence, together with our advances in understanding the mechanisms of the host's immune response to the bacterium due to genetic predisposition, have contributed to the understanding of its pathogenesis, transforming its clinical practice. Symptoms and development of the disease. It primarily affects the skin and may progress to a secondary stage, resulting in peripheral neuropathy and long-term disability and stigma. Damage to muscles, bones, skin, hair, nails, and mucous membranes is not directly related to the presence of Mycobacterium leprosy, but may be due to dystrophy caused by nerve damage. Leprosy is difficult to catch and has a long incubation period. This article has attempted to discuss the pathophysiological symptoms and treatment of leprosy. The purpose of this review is to inform the public about the complications of leprosy that can be prevented by taking preventive measures through leprosy awareness and to provide an insight into the herbal treatment of leprosy.

INTRODUCTION

Leprosy or Hansen's disease is a chronic bacterial infection caused by infection with Mycobacterium leprae (M. leprae). ^[1,2] Although nine-banded armadillos infect wildlife in the southern United States, M leprosy thrives on the feet of mice and is the primary method of breeding M leprosy in laboratories around the world. ^[3] Leprosy is common in tropical countries, especially underdeveloped and developing countries. In 1990, the World Health Organization (WHO) proposed a global goal of eradicating leprosy by the end of the 20th century. ^[4] Mycobacterium leprae is a chronic infectious disease known as leprosy or Hansen's disease, which was discovered centuries ago by the Norwegian physician Gerhard Hansen. It is considered an incurable disease. Brazil is known to have the second highest number of infections in the world. Nerve damage, skin deformities and progressive weakness are some of the symptoms of this disease. Worldwide. More than 5 million people are infected with mycobacteria, most of them in the United States. Asia. Pacific Islands and Africa. ^[5,6]



Fig.1 Indian women suffering from Leprosy

History of Leprosy

Leprosy has existed for many centuries, but its origins are unknown. ^[7] The disease has most likely spread around the world by human migrations such as relocation or colonization. ^[8] The origin of the term, leprosy, comes from biblical translations of Hebrew into Greek. The Hebrew word, "tsara'ath," translated as "leprosy" in the authorized version of the Old Testament, is a non-scientific term that indicates ritualistic defilement instead of a specific skin disease. ^[9] The Bible presents leprosy as a symbol that aggregates harmful consequences of impious behavior. ^[10] Due to these presentations, leprosy was commonly misunderstood as hereditary and incurable and was considered as "a divine punishment or curse," and this harmful image associated with the disease has resulted in inhumane treatments of patients such as leprosariums and mass executions. ^[7,11,12,13]

After the discovery of the bacillus M. Leprae as the cause of leprosy, countries set aims to eliminate leprosy. By providing MDT, an efficacious treatment based on the bacillus, to all government health facilities, the WHO successfully reduced the global prevalence of the disease. ^[14,15] However, complete elimination has yet to be reached as areas of high endemicity remain in many countries. For instance, in India, a country with more than 60% of the global burden of leprosy, the annual new case detection rate and prevalence rate per 10,000 people have remained nearly non-decreasing at 2.0 and 1.4, respectively, since 2007.^[16]

CLINICAL SIGNS AND SYMPTOMS

Bacterial leprosy targets the peripheral nervous system and causes many of the clinical symptom's characteristic of this mycobacterial infection.^[17] Lesions may affect the peripheral nerves of the skin, primarily the posterior tibial, ulnar, medial, and lateral peroneal nerves. Palpation of the nerve on physical examination reveals a superficial perineural osteofibrotic response. Nerve compression causes swelling, pain, and sensory and motor disturbances. Involvement of small cutaneous nerve fibers causes insomnia, dehydrated and impaired sensitivity to heat. In pure

TIJER || ISSN 2349-9249 || © April 2023 Volume 10, Issue 4 || www.tijer.org neurogenic leprosy, the neuropathy is asymmetric.^[18] This breed is common in India and Nepal. In other diseases, especially primary amyloidosis, and other hereditary diseases (e.g., Charcot-Marie-Tooth, Dejerin-Sottas, and Refsum diseases), peripheral nerve thickening should be considered in the differential diagnosis.^[19]

The musculoskeletal system is affected in 95% of cases. ^[20,21] Skeletal symptoms are often nonspecific because sensory loss after nerve injury leads to ulcers, deformities, and fractures. Osteoporosis is the second most common condition among people with leprosy.^[22] Acute orchitis, mainly associated with testicular lesions such as atrophy and erythema nodosum, has been reported in patients with leprosy. Eyes can cause direct infiltration or damage to the optic nerve. At the time of diagnosis, vision loss was documented in 11% of patients with polymycosis. The widespread variant described by Lucio and Alvarado in Mexico in 1851 is diffuse leprosy characterized by diffuse mucosal infiltration and atrophied appearance. The ears have a single protrusion. The main ocular symptoms of leprosy are blepharitis, keratitis, and entropion.^[23]



Fig.2 Mycobacterium leprae

ETIOLOGY

The etiology of M. leprae was identified in 1873 by the Norwegian physician Gerhard Armauer Hansen. That is why it is also called Hansen's wand. The classification, morphology, color, and biological characteristics of M. The scientific classification of leprosy is the class Schizomycetes, order Actinomycetales, family Mycobacteriaceae, and Mycobacterium. middle. Lefry - Straight or slightly curved, rounded at tip, 1.5-8 µm long, 0.2-0.5 µm in diameter. The stain is stained red with fuchsin by Ziehl-Nielsen (ZN) stain, and because of the high lipid content, the stain does not change even when washed with alcohol and acid, indicating the characteristics of ARB (acid-alcohol-fast bacilli). M. leprae differs from other mycobacteria in its arrangement, as it is arranged in parallel chains linked together to form spheres, like cigarettes in a box. When the Gram stain method is used, M. leprae is not visible on the Gram and appears as a negatively stained image or as a bead-like Gram-positive rod called a ghost. M. leprae primarily infects macrophages and Schwann cells. She was never raised in an artificial environment. Reproduction occurs by binary

fission and grows slowly (about 12-14 days) in the feet of mice. The temperature required for survival and reproduction is between 27^oC and 30^oC. This explains the greater frequency and less visceral involvement in superficial regions such as the skin, peripheral nerves, testes, and upper respiratory tract. M. leprae can survive 9 days in the environment. ^[24-28] Characteristics of the microstructure of M. leprae Microstructure of M. leprae Leprosy is often found in the Mycobacteriaceae. Electron microscopy showed that this bacillus contains cytoplasm, plasma membrane, cell wall and capsule. The cytoplasm contains structures common to Gram-positive microorganisms. The plasma membrane contains a permeable lipid bilayer with interacting proteins and protein surface antigens. The cell wall attached to the plasma membrane consists of peptidoglycan linked to a branched polysaccharide composed of arabinogalactan that supports mycosans and other mycobacterial-like lipoarabinomannan (LAM). The external structure of the capsule contains lipids, especially phthiocerol dimycocerol and phenol glycolipid (PGL-1), a trisaccharide linked to lipids by phenol molecules. This trisaccharide is a specific M. leprae antigen. ^[29,30] M. leprae genome Cole et al. 2001.22 Round. The estimated molecular weight is 2.2 x 109 Dalton's, 3,268,203 base pairs (bp), and the guanine + cytosine content is 57.8%. Compared to the Mycobacterium tuberculosis genome, it is 4,411,529 bp. blood. With a guanine + cytosine content of 65.6%, M. leprae appears to have undergone reductive evolution leading to a smaller genome enriched with inactive or completely deleted genes. It has 2770 genes with a coding rate of 49.5%, i.e 1604 protein-coding genes (M. leprae and M. tuberculosis) and 1116 (27%) pseudogenes. The latter are randomly distributed throughout the genome and may correspond to regulatory sequences or unrecognized residual gene mutations. This feature significantly reduces metabolic pathways, which explains why specific conditions are required for the growth of bacilli. ^[31,32] Tank M. Leprosy humans are reservoirs for M. leprae, but animals such as armadillos, chimpanzees and other great apes, soil, water, and some arthropods are natural reservoirs. ^[33-38]

MECHANISM OF LEPROSY TRANSMISSION

Transmission of leprosy is believed to occur through close and prolonged contact between a susceptible person and a person infected with the bacillus through inhalation of the bacilli present in nasal secretions or clear droplets. The primary route of transmission is through the nasal mucosa. ^[39-41] Transmission may occur primarily through skin erosion. ^[41,42] Other routes of transmission are possible, including blood, vertical transmission, breast milk, and insect bites. ^[43-45] It is believed that infected individuals may undergo a transient phase of nasal excretion of bacilli even if they do not develop disease. ^[46-49] M. Presence of leprosy on nasal swabs or biopsies. Seropositivity for specific bacillus antigens in healthy individuals living in endemic areas suggests a vector role in leprosy transmission. ^[46,47,50-60]



Fig.3 Patches on skin

TREATMENT

The World Health Organization (WHO) recommends age-adjusted multidrug therapy for the treatment of children, and these conditions are classified as minority balance and majority balance.^[61] Rifampicin, clofazimine, and dapsone (diaminodiphenylsulfone) have been used as first-line therapy. One paucibacillary case was treated with rifampicin, dapsone, and clofazimine for 6 months. The complex microbial condition was treated with rifampicin, dapsone and clofazimine for 12 months. All patients received this drug combination with monthly follow-up. In the United States, the regimen recommended by the National Hansen's Disease Program (NHDP) excludes clofazimine from the treatment of Parkinson's disease, so it is cheaper and has a longer treatment period. ^[62] Minocycline, ofloxacin, and clarithromycin are among the drugs used as second-line treatments. Advantages of multidrug therapy include prevention of dapsone resistance, rapid reduction of infection in infected individuals, and low rates of recurrence and response.^[63] However, long treatment periods and logistical difficulties make abstinence difficult to achieve. Leprosy patients have severe nerve damage, musculoskeletal disorders, and deformities, which can lead to discrimination in schools and difficulties in social life. Therefore, early diagnosis and treatment can reduce the prevalence and outcome of the disease in children. However, it is difficult for children to take medicine in the form of tablets and capsules, and they may not be able to chew the capsules, which can lead to incorrect dosage. Lack of oral solutions for children is a limiting factor in compliance.

PREVENTION

1. Prophylactic Immunity: The purpose of prophylactic immunity is to prevent infection, disease progression, or vaccination before or after exposure. Several vaccines have been shown to be effective, including Bacilli Calmette Guerin (BCG), LepVax, and Mycobacterium indicus pranii (MIP). ^[64] However, BCG is currently the only vaccine introduced to prevent leprosy. ^[65,66] A study was conducted on leprosy patients under 12 years of age attending a tertiary care hospital in eastern India. ^[67] The unvaccinated group had significantly more MB leprosy than the BCG-vaccinated group. This study demonstrates the role of BCG vaccine in enhancing cell-mediated immunity (CMI). In general, the protection of BCG vaccination against leprosy is estimated to be between 20% and 90%. ^[68, 69] However,

in countries with extensive BCG vaccine programs, leprosy is still common and, as with tuberculosis (TB), BCG vaccine protection against leprosy declines over time. ^[70] Also, in a study on the effect of BCG vaccination against leprosy from June 1987 to December 2006, BCG vaccination showed better protection against the MB strain than the PB strain. ^[71] However, the effectiveness of BCG vaccination is still controversial. Therefore, the development of more effective vaccines is very important. It can be used in addition to or instead of the BCG vaccine.

2. Chemoprophylaxis: Chemoprophylaxis using dapsone for leprosy was reported in the 1960s. ^[72] Studies have been conducted with dapsone/dapsone injection, rifampicin, and combinations of rifampicin, ofloxacin, and minocycline (ROM) for chemoprevention. A previous study showed that administration of a single dose of rifampicin (SDR) (25 mg/kg) to a relative of a new leprosy patient reduced the risk of developing clinical leprosy by 57% (95% CI 33–72). ^[73,74] Between 2015 and 2018, rifampicin post-exposure prophylaxis (SDR-PEP) was conducted in the Union Territory of Dadra and Nagar Haveli (DNH). ^[75] This study indicates that leprosy field research programs should focus on the health system. In addition, another study based on results from Bangladesh included in this study showed an additional protective effect of 80% (95% CI 50–92) of BCG plus rifampicin. ^[76] This finding highlights the potential of multimodal treatment strategies to reduce leprosy. RDS post-exposure prophylaxis was recommended by the WHO in 2018 and has been the preferred form of post-exposure prophylaxis for many years. BCG vaccine can prolong this. However, the extent to which SDR suppresses excess leprosy cases after BCG vaccination is difficult to assess because many cases occur before SDR intervention. ^[77,78] More research is needed on chemo preventive therapy to prevent leprosy. ^[79]

DIAGNOSIS

1. Serology and Molecular Diagnosis:

The importance of anti-PGL-I antibodies as a diagnostic serological test in the diagnosis of leprosy has been extensively studied. The disadvantage of this test is that it is not very sensitive to PB. ^[80] Another limitation of this experiment is that the individuals initially identified in the early stages of early disease and future disease are not family members. ^[81,82] Polymerase chain reaction (PCR) is very sensitive and unique, but requires high-quality laboratory equipment, so the quality is poor without research equipment and the quality is poor without equipment. Unused. ^[83,84] Finally, neutral antennas (ND) (ND-O-BSA) are considered useful for primary infection and relapse/control of M. leprae. ^[85] However, lack of funding and infrastructure limits the ability to deploy non-armed forces globally.

2. Slit-Skin Smear Test:

Bacilloscopic examination is an important method for an accurate diagnosis. Suitable sites for sampling are active lesions or lesions with altered sensation, the pavilion, and the contralateral elbow. ^[91] In the absence of trauma, intradermal shaving can be performed both at the tips of the ears and at the elbow. ^[90] The specificity of the photo smear is 100%, and the sensitivity is 50%. ^[91-93] Swabs of nasal mucosa, ears, forehead, chin, extensor surfaces of wrists, knees, body colds and/or skin lesions were the preferred sampling sites. After collection, acid-fast bacteria

(AFB) were examined using Fite stain or modified Ziel-Nielsen stain and Ridley's logarithmic scale or bacterial index (BI) was calculated. ^[87,94] A positive result indicates that the patient has MB. However, a negative result does not exclude a clinical diagnosis of leprosy and does not necessarily classify the patient as PD. The AFB staining method requires at least 10 organisms per gram of tissue for reliable detection under the microscope. The detection sensitivity of the organism is very low. [94] Microscopic examination detected positive bacilli in children (9.3%-25%). [86,88] Household contact is an important risk factor for infection in children. ^[95] The Cuban experience shows that 89% of diagnosed cases have at least one leper in the family. ^[96] Therefore, family history can be used as a diagnostic tool.

3. Skin Biopsy and Histological Examination: Skin biopsy is an important tool for diagnosing leprosy. The anterior edge of the most recently activated skin lesion was biopsied with the full thickness of the dermis, at least part of the subcutaneous fat lesion, and stained with the Fite-Faraco method. ^[97,98] Tissue samples were used for diagnosis. After extraction from body lesions, hematoxylin-eosin and Fite tissue stain were stained to examine the type, degree of invasion, signs of invasion, and AFB. Biopsy samples can be further analyzed for granuloma fraction, bacterial granuloma index (BIG) to assess AFB in the tissue and histopathological index. ^[83] BIG is a method used for the determination of AFB bacilli in a certain volume of tissue. Histological examination can help identify the type of leprosy and differentiate it from leprosy reactions. ^[83, 98] Histopathological data are used as the basis for the Ridley-Jopling spectral classification which defines five spectral types of leprosy (TL, BT, BB, BL, and LL). ^[81] Tuberculous poles are rare, and leprosy poles are filled with bacilli in inflammatory Virchow cell infiltrates. The specificity of skin biopsy specimens and histopathology ranged from 70% to 72%, but the sensitivity ranged from 49% to 70%. ^[99,100] The sensitivity and specificity of the WHO classification were 63% and 85%, respectively, using skin swabs and skin smears from 100 untreated, newly diagnosed leprosy patients who were classified as PB and MB according to the WHO classification.

4. Lepromin test: The lepromin test is based on the lepromin antigen (M. Leprosy caused by leprosy) and delayed hypersensitivity reaction (DTH) are read in two cycles. One is the early response (Fernandez) during the test and the other is the late response (Mitsuda). The Fernandez reaction was performed for 24 or 48 hours. Mitsuda's response was read after 21 days and showed resistance to Bacillus. Nodules >5 mm are benign. ^[101,102] TT/BT patients show a strong DTH skin response, whereas BL/LL patients do not develop a skin reaction to lepromin. ^[103] A previous study showed a difference in mean response size between non-contact and home contact tests using two soluble M. leprae antigens, suggesting that these antigens are not useful for leprosy diagnosis. ^[103] However, tests for lepromin (lepromin H and lepromin A) are useful for leprosy. Previously, skin test antigens for leprosy (lepromin A, Rees antigen and Convit antigen) have been used for about 40 years and have been shown to be safe when used in humans. ^[101] Recently, MLSA-LAM and MLCwA (M. leprae cell wall-associated antigen), obtained from M. leprae grown on armadillos. A clinical trial. ^[102] showed that both antigens at low doses had sensitivities of 20% and 25% but specificities of 100% and 95% in BT/TT leprosy patients. The sensitivity was 10% and 15% and the specificity was

70% and 60%, respectively, at high doses of both antigens, and an allergic reaction to leprosy antigen was observed in patients with BL/LL leprosy. ^[104] The primary leprosy skin test antigen (lepromin A) is generally safe for human use. The lepromin test is less accurate in diagnosing leprosy in children. The lepromin assay has several drawbacks, including inconsistent measurements due to mild DTH reactions in some individuals, titer differences between batches due to quality control issues, and lack of sufficient sensitivity and specificity. ^[102] These tests are still useful for validation for classification and prognostic purposes.

5. PCR testing: active surveillance and early detection of the disease represent the burden of leprosy and disability in society. ^[86] Polymerase chain reaction (PCR) is a molecular technique used to convert deoxyribonucleic acid (DNA) in M. lepra. leprous. A significant proportion of childhood leprosy cases remain AFB-negative on skin smears. ^[105] In these cases, additional methods are needed to confirm the diagnosis. Skin swab PCR is less invasive and less painful than in situ skin biopsy. ^[106] High sensitivity (87-100%) is observed in patients with PCR-positive types BI or LL. However, in patients with type BI or negative TT, PCR sensitivity may be low (30-83%). Over the past 30 years, PCR techniques have been developed to amplify various genetic targets in M. leprae. A PCR method was used to identify possible environments. ^[107-113]

MISCONCEPTIONS OF LEPROSY

A low level of knowledge about leprosy is associated with a high level of social exclusion. Instead of society's fears and stigma. Lack of knowledge about leprosy is common. It has been found to be related to negative attitudes towards lepers. ^[114-118] Misunderstanding between participants, such as that leprosy is transmitted by contact Stigma is growing in India in the current study. These misunderstandings are often associated with fear. Fear of disease and infection. ^[123,120121,122] Reduce the stigma of these misconceptions You need to solve, challenge, and increase your knowledge. this is also crucial Improving strategies for early detection of the disease, as lack of knowledge about leprosy is a major problem Factors contributing to late diagnosis. ^[124]

PERSON AFFECTED BY LEPROSY

We found that those affected were significantly more aware of leprosy than those closely related. Dignity and belonging to the community. This is like the findings of the Indian study. Lepers had higher educational scores than their relatives. ^[117] Affected individuals may be better educated because of their personal experiences with the disease. Because we often come across information about comfort and health When an employee is diagnosed. However, current knowledge of leprosy was low study. Like our results, several other studies in India reported low or insufficient levels. Knowledge of leprosy in affected persons. ^[125-127] The level of knowledge about leprosy may be low. It contributes to treatment discrepancies and needs to be addressed. ^[128] Health education should target the most stigmatized general public. in both countries. This can be done by targeting key influencers and authority figures. community such as village leaders who can influence others in the community Enable information filter.

TIJER || ISSN 2349-9249 || © April 2023 Volume 10, Issue 4 || www.tijer.org INTERVENTIONS TO IMPROVE LEPROSY KNOWLEDGE AND ACCEPTANCE

Our findings highlight the need for effective interventions that have a positive impact on acceptance. Increase your knowledge about leprosy and leprosy. We base our findings on regional differences. Gaps in knowledge, misunderstandings, beliefs, and fears are interference. adapted to specific cultures and contexts. ^[129,130] It is expected to be much more effective for cultivation. Positive attitudes and perceptions of people affected by leprosy compared to general messages. We believe that our knowledge indicates that certain topics should be prioritized in healthcare. Education in two countries: causes, mode of transmission, early symptoms, and contagiousness of leprosy. These results suggest that some messages are important but not essential. Now it is widespread. Knowing that leprosy could be cured was good for both. India and Indonesia. This probably reflects previous government announcements on education campaign. Knowledge gaps can be addressed through information, attitudes, beliefs, and fears. A complementary approach is needed. Education and awareness change are best combined. Health education states and behavior change interventions. ^[131,132]

INDIAN PLANTS USED IN THE TREATMENT OF LEPROSY

Sr. No	Botanic al Name	Family	Commo n name	Parts used	Photo	Reference
1.	Tinospo ra cordifoli a	Menisper maceae	Heart- leaved moonsee d	Stem		142
2.	Euphorb ia tirucalli	Euphorbia ceae	Firestick plants	Wood decocti ons		143

Table No.1 Indian Medicinal Plant

	<u>TIJER ISSN 2349-9249 © April 2023 Volume 10, Issue 4 www.tijer.org</u>						
3.	Calotrop is procera Linn.	Asclepiad aecae	Apple of Sodom	Differe nt parts of the plant Root		145,150	
4.	Cassia tora Linn.	Caesapini aceae	Sickle senna	Seeds		146	
5.	Sonchus arvensis Linn	Steraceae	Field milk thistle	Roots Leaves		151	
6.	Termina lia chebula Retz	Combreta ceae	Chebulie mvrobala n	Fruit		151	
TI,	JER230404(o 🛛 TIJER - 🛛	NTERNATIO	NAL RESE	EARCH JOURNAL www.tijer.org	339	

7.	Triumfet tapilosa Roth	Malvaceae	Burbark	Leaf Flower	151
8.	Dalbegi a sissoo Roxb	Fabaceae	Sisu North Indian Rosewoo d	Decoct ion of the bark and leaf Wood	151,152
9.	Musu paradisi ca	Musacaea	Banana	Astrin gent plant sap	153
10.	Mimosa pudica	Mimisace	Sensitive plant	Root	147
11.	Cordia dichoto ma (forsk)	Boraginac eae	Sebesten plum	Leaves Stem bark	154

12.	Pandanu stectoriu s Soland	Pandanacc ae	Screw pine, Ketki	Leaves	154
13.	Pterospe rmum acerifoli um Willd	Stericuliac eae	Karnikar a, Hathipail a	Stem bark	154
14.	Parkia biglobos a	Fabaceae	African locust bean	Dried leave	148
15.	Ocimum basilicu m L	Lamiaceae	Great basil	Dried stem bark	148

	<u>TIJER ISSN 2349-9249 © April 2023 Volume 10, Issue 4 www.tijer.org</u>							
16.	Mitracar pushirtu s L.	Rubiaceae	Girdlepo d	Root Plant		148		
17.	Bombax ceiba L.	Malvaceae	Semar	Roots of young plants Leaf Bark Flower		155,156		
18.	Ficus hispida Linn.f	Moraceae	Papasih	Fruits		155		
19.	Careaar borea Roxb	Leevthida ceac	Slow match tree	Leaves Bark		155		
20.	Taberna emontan a divaricat e L. Kurz.	Apocynac cae	Crepe gardenia	Leaves Seeds Roots		155		

21.	Dioscor ea transver sa R. Br.	Dioscorea ceae	Pencil yam, Pokmaso	Seeds Tuber	155
22.	Bauhini a variegat a Linn.	Fabaceae	Orchid tree	Plant Bark	149
23.	Striga hermont hica	Orobanch aceae	Purple witchwe ed	Plant	157
24.	Anacard ium occident ale L.	Anacardia ceae	Cashew	Leaves	158

	<u>TIJ</u>	ER ISSN 23	<u>49-9249 ©</u>	April 2023	<u>3 Volume 10, Issue 4 www.tijer.org</u>	L
25.	Afzelia africana Sm.	Caesalpini aceae	Afzelia	Bark powde r		150
26.	Vitexdo niana Sweet	Lamiaceae	Nkokoro	Root		159
27.	Albizzia libbeck	Mimosace ae	Lebbek treeor flea tree	Seed oil Bark		160
28.	Amarant hus spinosis	Amaranth aceae	Prickly amaranth	Whole plant		161
29.	Gmelina arborea Roxb.	Verbencea e	Goomar treak	Fruits Flower s		144

30.	Centella asiatica Linn	Apiaceous	Asiatic pennywo rt	Whole plant	161,162
31.	Hiptage benghal ensis (L.)	Malpighia ceae	Hiptage	Leaves Bark	161
32.	Holarrhe na antidvse nterica Foxh	Apocynac eae	Bitter Oleander	Bark Seeds	161

TIJER || ISSN 2349-9249 || © April 2023 Volume 10, Issue 4 || www.tijer.org SYNTHETIC DRUGS USED IN LEPROSY

Sr.No	Synthetic	Activity	Structure	Reference
	drugs used	·		
1	Clofazimine	Binds to mycobacterium DNA, inhibits mycobacterium growth 16B		163,164
2	Dapsone	Inherent level of bactericidal activity 15B		165,166
3	Rifampicin	High bactericidal activity on M. leprae 1		167,168
4	Ofloxacin	Inhibit bacterial DNA gyrase 18B		169,170
5	Prednisolone	Inhibition of macrophage accumulation		169,171

Table No. 2 Synthetic Drugs

CONCLUSION

Leprosy, also known as Abhansen's disease, is a chronic bacterial infection caused by the Lepra bacterium. The disease mainly affects the skin, peripheral nerves and mucous membranes of the upper respiratory tract and causes several symptoms and complications. Diagnosis of leprosy is based on clinical signs and symptoms, as well as laboratory tests to determine the presence of bacteria. Early detection and treatment are essential to prevent serious disorders and deformities. Treatment for leprosy includes a combination of antibiotics that target the bacteria along with supportive

treatment and rehabilitation to manage complications and disability. Stigma and discrimination against people with leprosy remain a major problem, underscoring the importance of raising awareness of the disease and dispelling misconceptions. Overall, while significant progress has been made in the prevention, diagnosis, and treatment of leprosy, much remains to be done to eradicate the disease and ensure that those affected receive the treatment and support they need.

RESULT

This review article provides an overview of leprosy, a chronic infectious disease caused by Mycobacterium leprae, and its treatment. The epidemiology, transmission, clinic, diagnosis, and treatment of leprosy are discussed. They emphasize the importance of early diagnosis and treatment to prevent disease-induced disability and deformity. We also discuss the challenges of leprosy management and the need for a multidisciplinary approach involving health authorities, providers, and communities. Overall, the review articles provide a comprehensive understanding of leprosy, treatment and herbal treatments for leprosy that can benefit people and highlight the need for more awareness and research to eradicate the disease.

Reference

1. S. Ghosh and S. Chaudhuri, "Chronicles of Gerhard Henrik Armauer Hansen's life and work," Indian Journal of Der- matology, vol. 60, no. 3, p. 219, 2015.

2. W. J. Britton and D. N. Lockwood, "Leprosy," The Lancet, vol. 363, no. 9416, pp. 1209–1219, 2004.

3. C. F. Paige, R. W. Truman, and D. T. Scholl, "Prevalence and incidence density of Mycobacterium leprae and Trypanosoma cruzi infections within a population of wild nine-banded armadillos," The American Journal of Tropical Medicine and Hygiene, vol. 67, no. 5, pp. 528–532, 2002.

4. World Health Organization (WHO), "Leprosy," 2022, https://www.who.int/en/news-room/fact-sheets/detail/ leprosy

5. Barbosa-filho JM et al. Natural products with antileproticability of activity. Brazilian J Pharmacogn. 17 (1): 2007: 141-148.

6. Gupta A et al. Antileprotic Potential of Ethnomedicinal Herbs: Aure. Thus Review. Drug Invent Today. 2 (3); 2010: 191-193.

7. Trautman JR. A brief history of hansen's disease. Bull N Y Acad Med (1984) 60(7):689–95.

8. Monot M, Honoré N, Garnier T, Araoz R, Coppée J-Y, Lacroix C, et al. On the origin of leprosy. Science (2005) 308(5724):1040–2. doi: 10.1126/science/1109759

9. Browne SG. Some aspects of the history of leprosy: the leprosie of yesterday. Proc R Soc Med (1975) 68(8):485–
93. doi: 10.1177/003591577506800809

TIJER || ISSN 2349-9249 || © April 2023 Volume 10, Issue 4 || www.tijer.org 10. Lewis G. A lesson from Leviticus: Leprosy. Man (Lond) (1987) 22(4):593. doi: 10.2307/2803354

11. Grzybowski A, Sak J, Pawlikowski J, Nita M. Leprosy: Social implications from antiquity to the present. Clin Dermatol (2016) 34(1):8–10. doi: 10.1016/ j. clindermatol.2015.10.009

12. Robertson J. The leprosy asylum in India: 1886-1947. J Hist Med Allied Sci (2009) 64(4):474-517. doi: 10.1093/jhmas/jrp014

13. Couto Dal Secco RG, França K, Castillo D, AlHarbi M, Lotti T, Fioranelli M, et al. A synopsis of the history of hansen's disease. Wien Med Wochenschr (2017) 167(Suppl 1):27–30. doi: 10.1007/s10354-017-0590-2

14. Manglani PR, Arif MA. Multidrug therapy in leprosy. J Indian Med Assoc (2006) 104(12):686-

15. Smith CS, Aerts A, Saunderson P, Kawuma J, Kita E, Virmond M. Multidrug therapy for leprosy: A game changer on the path to elimination. Lancet Infect Dis (2017) 17(9): e293–7. doi: 10.1016/S1473-3099(17)30418-8

Mohanty P, Naaz F, Bansal A, Kumar D, Gupta U. Challenges beyond elimination in leprosy. Int J Mycobacteriol (2017) 6(3):222. doi: 10.4103/ ijmy. ijmy_70_17

17. Garg R, Dehran M. Leprosy: a precipitating factor for complex regional pain syndrome. Minerva Anestesiol. 2010; 76:758-60.

18. Sehgal V, Sardana K, Dogra S. Management of complicationsfollowing leprosy: an evolving scenario. J Dermatolog Treat.2007;18:366-74.

19. Maki DD, Yousem DM, Corcoran C, Galetta SL. MR imag-ing of Dejerine-Sottas disease. AJNR Am J Neuroradiol.1999;20:378-80.

20. Faget GH, Mayoral A. Bone changes in leprosy, a clinical androentgenological study of 505 cases. Radiology. 1944; 42:1-13.

21. Chhabriya BD, Sharma NC, Bansal NK, Agrawal GR. Bone changes in leprosy. A study of 50 cases. Indian J Lepr. 1985; 57:632-9.

22. Martínez de Lagrán Z, Arrieta-Egurrola A, González-Pérez R,

Soloeta-Arechavala R. Complicaciones seas en un paciente con lepra lepromatosa. Actas Dermosifiliogr. 2009; 100:615-7.

23. Vargas-Ocampo F. Diffuse leprosy of Lucio and Latapi: a histo-logic study. Lepr Rev. 2007; 78:248-60

24. Rees RJW, Young DB. The microbiology of leprosy. In: Hastings RC, editor. Leprosy. 2nd ed. New York: Churchill Livingstone; 1994. p.49-83.

25. Nolte FS, Metchok B. Mycobacterium. In: Murray PR, Baron EJ, Pfaller MA, Tenover FC, Yolken RH, editors. Manual of clinical microbiology. 6th ed. Washington: American Society for Microbiology; 1995. p. 400-37.

26. Shepard CC. Temperature optimum of Mycobacterium leprae in mice. J Bacteriol. 1965; 90:1271-5.

27. Hastings RC, Brand PW, Mansfield RE, Ebner JD. Bacterial density in the skin in lepromatous leprosy as related to temperature. Lepr Rev. 1968;39(2):71-4.

28. Desikan KV. Viability of Mycobacterium leprae outside the human body. Lepr Rev. 1977; 48:231-5.

29. Draper P. The bacteriology of Mycobacterium. Tubercle. 1983; 64:43-56.

30. Hirata T. Electron microscopic observations of cell wall and cytoplasmic membrane in murine and human leprosy bacilli. Int J Lepr Other Mycobact Dis. 1985; 53:433-40.

31. Cole ST, Eiglmeier K, Parkhill J, James KD, Thomson NR, Wheeler PR, et al. massive gene decay in the leprosy bacillus. Nature. 2001; 409:1007-11.

32. Vissa VD, Brennan PJ. The genome of Mycobacterium leprae: a minimal mycobac- terial gene set. Genome Biol. 2001;2: REVIEWS1023.

33. Deps PD, Antunes JM, Tomimori-Yamashita J. Detection of Mycobacterium leprae infection in wild nine-banded armadillos (Dasypus novemcinctus) using the rapid ML Flow test. Rev Soc Bras Med Trop. 2007; 40:86-7.

34. Donham KJ, Leininger JR. Spontaneous leprosy-like disease in a chimpanzee. J Infect Dis. 1977; 136:132-6.

35. Walsh GP, Meyers WM, Binford CH, Gerone PJ, Wolf RH, Leininger JR. Leprosy-- a zoonosis. Lepr Rev. 1981; 52:77-83.

36. Kazda J, Ganapati R, Revankar C, Buchanan TM, Young DB, Irgens LM. Isolation of environment-derived Mycobacterium leprae from soil in Bombay. Lepr Rev. 1986; 57:201-8.

37. Matsuoka M, Izumi S, Budiawan T, Nakata N, Saeki K. Mycobacterium leprae DNA in daily using water as a possible source of leprosy infection. Indian J Lepr. 1999; 71:61-7.

38. Bona SH, Fonseca APM, Silva ACL, Costa RJ. Bacilos álcool-ácido resistentes no Culex fatigans. A Bras Dermatol. 1985; 60:163-70.

39. Shepard CC. The nasal excretion of Mycobacterium leprae in leprosy. Int J Lepr. 1962; 30:10-8.

40. Martins AC, Miranda A, Oliveira ML, Bührer-Sékula S, Martinez A. Estudo da mucosa nasal de contatos de hanseníase, com positividade para o antígeno glico- lipídio fenólico 1. Braz J Otorhinolaryngol. 2010; 76:579-87.

41. Job CK. Nasal mucosa and abraded skin are the two routes of entry of Mycobacterium leprae. Star. 1990; 49:1.

42. Ghorpade A. Inoculation (tattoo) leprosy: a report of 31 cases. J Eur Acad Dermatol Venereol. 2002; 16:494-9.

43. Santos AR, Balassiano V, Oliveira ML, Pereira MA, Santos PB, Degrave WM, et al. Detection of Mycobacterium leprae DNA by polymerase chain reaction in the blood of individuals, eight years after completion of anti-leprosy therapy. Mem Inst Oswaldo Cruz. 2001; 96:1129-33.

44. Melsom R, Harboe M, Duncan ME, Bergsvik H. IgA and IgM antibodies against Mycobacterium leprae in cord sera and in patients with leprosy: an indicator of intrauterine infection in leprosy. Scand J Immunol. 1981; 14:343-52.
45. JC. The presence of M. leprae in human milk. Lepr Rev. 1967; 38:239-42.

46. Klatser PR, van Beers S, Madjid B, Day R, de Wit MY. Detection of Mycobacterium leprae nasal carriers in populations for which leprosy is endemic. J Clin Microbiol.1993;31:2947-51.

47. Hatta M, van Beers SM, Madjid B, Djumadi A, de Wit MY, Klatser PR. Distribution and persistence of Mycobacterium leprae nasal carriage among a population in which leprosy is endemic in Indonesia. Trans R Soc Trop Med Hyg. 1995; 89:381- 5.

48. van Beers SM, de Wit MY, Klatser PR. The epidemiology of Mycobacterium leprae: recent insight. FEMS Microbiol Lett. 1996; 136:221-30.

49. Cree IA, Smith WC. Leprosy transmission and mucosal immunity: towards eradi- cation Lepr Rev. 1998; 69:112-21.

50. Pattyn SR, Ursi D, Ieven M, Grillone S, Raes V. Detection of Mycobacterium leprae by the polymerase chain reaction in nasal swabs of leprosy patients and their con- tacts. Int J Lepr Other Mycobact Dis. 1993; 61:389-93.

51. van Beers SM, Izumi S, Madjid B, Maeda Y, Day R, Klatser PR. An epidemiological study of leprosy infection by serology and polymerase chain reaction. Int J Lepr Other Mycobact Dis. 1994; 62:1-9.

52. Ramaprasad P, Fernando A, Madhale S, Rao JR, Edward VK, Samson PD, et al. Transmission and protection in leprosy: indications of the role of mucosal immu- nity. Lepr Rev. 1997; 68:301-15.

53. Izumi S, Budiawan T, Saeki K, Matsuoka M, Kawatsu K. An epidemiological study on Mycobacterium leprae infection and prevalence of leprosy in endemic villages by molecular biological technique. Indian J Lepr. 1999; 71:37-43.

54. Patrocínio LG, Goulart IM, Goulart LR, Patrocínio JA, Ferreira FR, Fleury RN. Detection of Mycobacterium leprae in nasal mucosa biopsies by the polymerase chain reaction. FEMS Immunol Med Microbiol. 2005; 44:311-6.

55. Job CK, Jayakumar J, Kearney M, Gillis TP. Transmission of leprosy: a study of skin and nasal secretions of household contacts of leprosy patients using PCR. Am J Trop Med Hyg. 2008; 78:518-21.

56. Baumgart KW, Britton WJ, Mullins RJ, Basten A, Barnetson RS. Subclinical infec- tion with Mycobacterium leprae--a problem for leprosy control strategies. Trans R Soc Trop Med Hyg. 1993; 87:412-5.

57. Brasil MTLRF, Oliveira LR, Melo CS, Nakamura PM, Rimoli NS, Cavalari FS et al. Aplicação do teste Elisa anti-PGL-1 em localidade com alta edemicidade de han- seníase, na Região Norte do Estado de São Paulo. Hansen Int. 1998; 23:35-48.

58. de Wit MY, Douglas JT, McFadden J, Klatser PR. Polymerase chain reaction for detection of Mycobacterium leprae in nasal swab specimens. J Clin Microbiol. 1993; 31:502-6.

59. Fine PE, Sterne JA, Pönnighaus JM, Bliss L, Saui J, Chihana A, et al. Household and dwelling contact as risk factors for leprosy in northern Malawi. Am J Epidemiol. 1997; 146:91-102.

60. Moet FJ, Meima A, Oskam L, Richardus JH. Risk factors for the development of clinical leprosy among contacts, and their relevance for targeted interventions. Lepr Rev. 2004; 75:31026.

61.World Health Organization, "Guidelines for the diagnosis, treatment and prevention of leprosy," 2022, https://apps. who.int/iris/bitstream/handle/10665/274127/ 9789290226383-eng.

62.National Hansen's Disease Program, "Health resources and Services administration, "recommended treatment regimens"," 2022, https://www.hrsa.gov/hansens-disease/ diagnosis/recommended-treatment.html.

63.W. Alemu Belachew and B. Naafs, "Position statement: leprosy: diagnosis, treatment and follow-up," Journal of the European Academy of Dermatology and Venereology, vol. 33, no. 7, pp. 1205–1213, 2019.

64. World Health Organization, "BCG vaccine: WHO position paper, February 2018 – recommendations," Vaccine, vol. 36, no. 24, pp. 3408–3410, 2018.

65. M. S. Duthie, P. Saunderson, and S. G. Reed, "The potential for vaccination in leprosy elimination: new tools for targeted interventions," Memorias do Instituto Oswaldo Cruz, vol. 107, no. suppl 1, pp. 190–196, 2012.

66. M. S. Duthie, T. P. Gillis, and S. G. Reed, "Advances and hurdles on the way toward a leprosy vaccine," Human Vaccines, vol. 7, no. 11, pp. 1172–1183, 2011.

67. S. Sarkar, T. Sarkar, A. C. Patra et al., "BCG vaccination: effects on the patterns of pediatric leprosy," Journal of Family Medicine and Primary Care, vol. 9, no. 7, p. 3673, 2020.

68. C. S. Merle, S. S. Cunha, and L. C. Rodrigues, "BCG vac- cination and leprosy protection: review of current evidence and status of BCG in leprosy control," Expert Review of Vaccines, vol. 9, no. 2, pp. 209–222, 2010.

69. R. Richardus, A. vanHooij, S.J.F.vandenEedenetal., "BCG and adverse events in the context of leprosy," Frontiers in Immunology, vol. 9, p. 629, 2018.

70. H. J. de Matos, D. J. Blok, S. J. de Vlas, and J. H. Richardus, "Leprosy new case detection trends and the future effect of preventive interventions in para['] state, Brazil: a modelling study," PLoS Neglected Tropical Diseases, vol. 10, no. 3, Article ID e0004507, 2016.

71. N. C. Du ppre, L. A. Camacho, S. S. da Cunha et al., "Ef- fectiveness of BCG vaccination among leprosy contacts: a cohort study," Transactions of the Royal Society of Tropical Medicine and Hygiene, vol. 102, no. 7, pp. 631–638, 2008.

72. A. Schoenmakers, L. Mieras, T. Budiawan, and W. H. van Brakel, "The state of affairs in post-exposure leprosy prevention: a descriptive meta-analysis on immuno- and chemo-prophylaxis," Research and Reports in Tropical Medicine, vol. 11, pp. 97–117, 2020.

73. F. J. Moet, L. Oskam, R. Faber, D. Pahan, and J. H. Richardus, "A study on transmission and a trial of chemoprophylaxis in contacts of leprosy patients: design, methodology and re- cruitment findings of COLEP," Leprosy Review, vol. 75, no. 4, pp. 376–388, 2004.

74. L. N. Nguyen, J. L. Cartel, and J. H. Grosset, "Chemopro- phylaxis of leprosy in the Southern Marquesas with a single 25 mg/kg dose of rifampicin. results after 10 years," Leprosy Review, vol. 71, pp. S33–S35, 2000.

75. A. Tiwari, L. Mieras, K. Dhakal et al., "Introducing leprosy post-exposure prophylaxis into the health systems of India, Nepal and Indonesia: a case study," BMC Health Services Research, vol. 17, no. 1, p. 684, 2017.

76. R. P. Schuring, J. H. Richardus, D. Pahan, and L. Oskam, "Protective effect of the combination BCG vaccination and rifampicin prophylaxis in leprosy prevention," Vaccine, vol. 27, no. 50, pp. 7125–7128, 2009.

77. R. Richardus, K. Alam, K. Kundu et al., "Effectiveness of single-dose rifampicin after BCG vaccination to prevent leprosy in close contacts of patients with newly diagnosed leprosy: a cluster randomized controlled trial," International Journal of Infectious Diseases, vol. 88, pp. 65–72, 2019.

78. R. A. Richardus, K. Alam, D. Pahan, S. G. Feenstra, A. Geluk, and J. H. Richardus, "The combined effect of chemopro- phylaxis with single dose rifampicin and immunoprophy-laxis with BCG to prevent leprosy in contacts of newly diagnosed leprosy cases: a cluster randomized controlled trial (MALTALEP study)," BMC Infectious Diseases, vol. 13, no. 1, p. 456, 2013.79. A. Tiwari, D. J. Blok, M. Arif, and J. H. Richardus, "Leprosy post-exposure prophylaxis in the Indian health system: a cost-effectiveness analysis," PLoS Neglected Tropical Diseases, vol. 14, no. 8, Article ID e0008521, 2020

80. Cho SN, Cellona RV, Villahermosa LG, Fajardo TT Jr, Balagon MV, Abalos RM, et al. Detection of phenolic glycolipid I of

Mycobacterium leprae in sera from leprosy patients before and after start of multidrug therapy. Clin Diagn Lab Immunol 2001; 8:138-42.

81. Soares DJ, Failbus S, Chalise Y, Kathet B. The role of IgM antiphenolic glycolipid-1 antibodies in assessing household contacts of leprosy patients in a low endemic area. Lepr Rev 1994; 65:300-4.

82. Kampirapap K. Assessment of subclinical leprosy infection through the measurement of PGL-1 antibody levels in residents of a former leprosy colony in Thailand. Lepr Rev 2008; 79:315-9.

83. Banerjee S, Ray D, Bandyopadhyay D, Gupta S, Gupta S, Ghosal C, et al. Development and application of a new efficient and sensitive multiplex polymerase chain reaction (PCR) in diagnosis of leprosy. J Indian Med Assoc 2008; 106:436-40.

84. Shetty VP, Doshi RP. Detection and classification of leprosy: Future needs and strategies. Indian J Lepr 2008; 80:139-47.

85. Wu Q, Yin Y, Zhang L, Chen X, Yu Y, Li Z, et al. A study on a possibility of predicting early relapse in leprosy using a ND-O-BSA based ELISA. Int J Lepr Other Mycobact Dis 2002; 70:1-8.

86. S. V. Gitte, N. S. Ramanath, and K. M. Kamble, "Childhood leprosy in an endemic area of Central India," Indian Pedi- atrics, vol. 53, no. 3, pp. 221–224, 2016.

87. K. Eichelmann, S. E. Gonza'lez Gonza'lez, J. C. Salas-Alanis, and J. Ocampo-Candiani, "Leprosy. an update: definition, pathogenesis, classification, diagnosis, and treatment," Actas Dermo-Sifiliogra'ficas, vol. 104, no. 7, pp. 554–563, 2013.

88. A. G. Rao, "Study of leprosy in children," Indian Journal of Leprosy, vol. 81, pp. 195–197, 2009.

89. I. M. Romero-Montoya, J. C. Beltra n-Alzate, D. C. Ortiz- Mar in, A. Diaz-Diaz, and N. Cardona-Castro, "Leprosy in Colombian children and adolescents," The Pediatric Infec- tious Disease Journal, vol. 33, no. 3, pp. 321-322, 2014.

90. S. C. Moreira, C. J. D. C. Batos, and L. Tawil, "Epidemio- logical situation of leprosy in Salvador from 2001 to 2009," Anais Brasileiros de Dermatologia, vol. 89, no. 1, pp. 107–117, 2014.

91. E. E. V. Quilter, C. R. Butlin, S. Singh, K. Alam, and D. N. J. Lockwood, "Patients with skin smear positive leprosy in Bangladesh are the main risk factor for leprosy devel- opment: 21-year follow-up in the household contact study (COCOA)," PLoS Neglected Tropical Diseases, vol. 14, no. 10, Article ID e0008687, 2020.

92. S. Banerjee, N. Biswas, N. Kanti Das et al., "Diagnosing leprosy: revisiting the role of the slit-skin smear with critical analysis of the applicability of polymerase chain reaction in diagnosis," International Journal of Dermatology, vol. 50, no. 12, pp. 1522–1527, 2011.

93. M. D. C. S. Azevedo, N. M. Ramuno, L. R. V. Fachin et al., "qPCR detection of Mycobacterium leprae in biopsies and slit skin smear of different leprosy clinical forms," Brazilian Journal of Infectious Diseases, vol. 21, no. 1, pp. 71–78, 2017.

94. I. Bhat, J. Madhukara, P. Rout, J. Elizabeth, and S. Kumaran, "Comparison of bacillary index on slit skin smear with bacillary index of granuloma in leprosy and its relevance to present therapeutic regimens," Indian Journal of Derma- tology, vol. 60, no. 1, p. 51, 2015.

95. S. Jain, R. G. Reddy, S. N. Osmani, D. N. J. Lockwood, and S. Suneetha, "Childhood leprosy in an urban clinic, Hyderabad, India: clinical presentation and the role of household contacts," Leprosy Review, vol. 73, no. 3, pp. 248–253, 2002.

96. J. L. Ruiz-Fuentes, R. Rumbaut Castillo, L. D. L. C. Hurtado Gasco[']n, and F. Pastrana, "Leprosy in children: a Cuban experience on leprosy control," BMJ Paediatrics Open, vol. 3, no. 1, Article ID e000500, 2019.

97. C. Massone, W. A. Belachew, and A. Schettini, "Histopa- thology of the lepromatous skin biopsy," Clinics in Dermatology, vol. 33, no. 1, pp. 38–45, 2015.

98. A. N. Martinez, C. F. P. C. Britto, J. A. C. Nery et al., "Evaluation of real-time and conventional PCR targeting complex 85 genes for detection of Mycobacterium leprae DNA in skin biopsy samples from patients diagnosed with leprosy," Journal of Clinical Microbiology, vol. 44, no. 9, pp. 3154–3159, 2006.

99. W. van Brakel, H. Cross, E. Declercq et al., "Review of leprosy research evidence (2002–2009) and implications for current policy and practice," Leprosy Review, vol. 81, no. 3, pp. 228–275, 2010.

100. D. N. J. Lockwood, P. Nicholls, W. C. S. Smith et al., "Comparing the clinical and histological diagnosis of leprosy and leprosy reactions in the INFIR cohort of Indian patients with multibacillary leprosy," PLoS Neglected Tropical Dis- eases, vol. 6, no. 6, Article ID e1702, 2012.

101. K. Ro'ltgen, G. Pluschke, J. S. Spencer, P. J. Brennan, and C. Avanzi, "The immunology of other mycobacteria: M. ulcerans, M. leprae," Seminars in Immunopathology, vol. 42, no. 3, pp. 333–353, 2020.

102. B. L. Rivoire, N. A. Groathouse, S. TerLouw et al., "Safety and efficacy assessment of two new leprosy skin test antigens: randomized double blind clinical study," PLoS Neglected Tropical Diseases, vol. 8, no. 5, Article ID e2811, 2014.

103. U. Sengupta, "Recent laboratory advances in diagnostics and monitoring response to treatment in leprosy," Indian Der- matology Online Journal, vol. 10, no. 2, p. 106, 2019.

104. P. Gurung, C. M. Gomes, S. Vernal, and M. M. G. Leeflang, "Diagnostic accuracy of tests for leprosy: a systematic review and meta-analysis," Clinical Microbiology and Infections, vol. 25, no. 11, pp. 1315–1327, 2019.

105. A. K. Simon, G. A. Hollander, and A. McMichael, "Evolution of the immune system in humans from infancy to old age," Proceedings of the Royal Society B: Biological Sciences, vol. 282, no. 1821, Article ID 20143085, 2015.

106. R. Kamal, M. Natrajan, K. Katoch, and V. M. Katoch, "Evaluation of diagnostic role of in situ PCR on slit-skin smears in pediatric leprosy," Indian Journal of Leprosy, vol. 82, no. 4, pp. 195–200, 2010.

107. A. N. Martinez, C. Talhari, M. O. Moraes, and S. Talhari, "PCR-based techniques for leprosy diagnosis: from the laboratory to the clinic," PLoS Neglected Tropical Diseases, vol. 8, no. 4, Article ID e2655, 2014

108. A. N. Martinez, M. Ribeiro-Alves, E. N. Sarno, and M. O. Moraes, "Evaluation of qPCR-based assays for leprosy diagnosis directly in clinical specimens," PLoS Neglected Tropical Diseases, vol. 5, no. 10, Article ID e1354, 2011.

109. R. S. Gama, L. A. Leite, L. T. Colombo, and L. A. D. O. Fraga, "Prospects for new leprosy diagnostic tools, a narrative re- view considering ELISA and PCR assays," Revista da Sociedade Brasileira de Medicina Tropical, vol. 53, Article ID e20200197, 2020.

110. V. Singh, R. P. Turankar, and A. Goel, "Real-time PCR-based quantitation of viable Mycobacterium leprae strain from clinical samples and environmental sources and its genotype in multi-case leprosy families of India," European Journal of Clinical Microbiology & Infectious Diseases, vol. 39, no. 11, pp. 2045–2055, 2020.

111. L. E[´]. C. Marques, C. C. Frota, J. D. S. Quetz et al., "Evaluation of 16S rRNA qPCR for detection of Mycobacterium leprae DNA in nasal secretion and skin biopsy samples from multibacillary and paucibacillary leprosy cases," Pathogens and Global Health, vol. 112, no. 2, pp. 72–78, 2018.

112. R. S. Gama, T. A. R. Gomides, C. F. M. Gama et al., "High frequency of M. leprae DNA detection in asymptomatic household contacts," BMC Infectious Diseases, vol. 18, no. 1, p. 153, 2018.113. R. S. Gama, M. L. M. D. Souza, E. N. Sarno et al., "A novel integrated molecular and serological analysis method to predict new cases of leprosy amongst household contacts," PLoS Neglected Tropical Diseases, vol. 13, no. 6, Article ID e0007400, 2019.

114. Adhikari B, Shrestha K, Kaehler N, Raut S, Chapman SR. Community attitudes towards leprosy affected persons in Pokhara municipality of western Nepal. J Nepal Health Res Counc 2014.

115. O'Donoghue J, Ishengoma A, Masao H, Mbega M. Evaluation of a sustained 7-year health education campaign on leprosy in Rufiji District, Tanzania. Lepr Rev 1998; 69:57–74. https://doi.org/10.5935/0305-7518.19980007 PMID: 9628096

116. Singh R, Singh B, Mahato S. Community knowledge, attitude, and perceived stigma of leprosy amongst community members living in Dhanusha and Parsa districts of Southern Central Nepal. PLoS Negl Trop Dis 2019; 13: e0007075. https://doi.org/10.1371/journal.pntd.0007075 PMID: 30633780

117. Seshadri D, Khaitan BK, Khanna N, Sagar R. The tangled web: a study of knowledge and attitude towards leprosy from a tertiary care hospital in India. Indian J Lepr 2014:27–41. PMID: 25591277

118. van't Noordende AT, Korfage I, Lisam S, Arif MA, Kumar A, van Brakel WH. The role of perceptions and knowledge of leprosy in the elimination of leprosy: A baseline study in Fatehpur district, northern India. PLoS Negl Trop Dis 2019; 13: e0007302. <u>https://doi.org/10.1371/journal.pntd.0007302</u>

PMID: 30951526

119. Ebenso B, Newell J, Emmel N, Adeyemi G, Ola B. Changing stigmatisation of leprosy: an exploratory,

qualitative life course study in Western Nigeria. BMJ Glob Heal 2019; 4: e001250. https://doi.org/10.1136/bmjgh-2018-001250 PMID: 30997168

120. Sottie CA, Darkey J. Living with stigma: Voices from the Cured Lepers' village in Ghana. Soc Work Health Care 2019; 58:151–65. https://doi.org/10.1080/00981389.2018.1526842 PMID: 30321131

121. Hofstraat K, van Brakel WH. Social stigma towards neglected tropical diseases: a systematic review. In Health 2016; 8: i53–70. https://doi.org/10.1093/inthealth/ihv071 PMID: 26940310

122. de Stigter DH, Geus LD, Heynders ML. Leprosy: between acceptance and segregation. Community behaviour towards persons affected by leprosy in eastern Nepal. Lepr Rev 2000; 71:492–8. https://doi.org/10.5935/0305-7518.20000051 PMID: 11201904

123. Sermrittirong S, Van Brakel W. Stigma in leprosy: concepts, causes and determinants. Lepr Rev 2014; 85:36–47. PMID: 24974441

124. Lasto´ria JC, Abreu MAMM de. Leprosy: review of the epidemiological, clinical, and etiopathogenicaspectspart 1. A Bras Dermatol 2014; 89:205–18. https://doi.org/10.1590/abd1806-4841.20142450PMID: 24770495

125. Grewal I, Negi Y, Kishore J, Adhish S V. Knowledge, and attitude about Leprosy in Delhi in post elimination phase. Indian J Lepr 2013; 85. PMID: 24724234

126. Barkataki P, Kumar S, Rao PS. Knowledge of and attitudes to leprosy among patients and community members: a comparative study in Uttar Pradesh, India. Lepr Rev 2006; 77:62–8. PMID: 16715691

127. Mankar MJ, Joshi SM, Velankar DH, Mhatre RK, Nalgundwar AN. A comparative study of the quality of life, knowledge, attitude, and belief about leprosy disease among leprosy patients and community members in Shantivan Leprosy Rehabilitation centre, Nere, Maharashtra, India. J Glob Infect Dis 2011; 3:378. https://doi.org/10.4103/0974-777X.91063 PMID: 22224003

128. Girão RJS, Soares NLR, Pinheiro JV, da Paz Oliveira G, De Carvalho SMF, De Abreu LC, et al. Leprosy treatment dropout: A sistematic review. Int Arch Med 2013; 6:34. <u>https://doi.org/10.1186/1755-7682-6-</u> 34 PMID: 24000954

129. Opala J, Boillot F. Leprosy among the Limba: illness and healing in the context of world view. Soc SciMed 1996; 42:3–19. https://doi.org/10.1016/0277-9536(95)00026-7 PMID: 8745104

130. Chen PC, Sim HC. The development of culture-specific health education packages to increase case-finding of leprosy in Sarawak. Southeast Asian J Trop Med Public Health 1986; 17:427–32. PMID:3563610

131. Seligman HK, Wallace AS, DeWalt DA, Schillinger D, Arnold CL, Shilliday BB, et al. Facilitating behavior change with low-literacy patient education materials. Am J Health Behav 2007; 31: S69–78. https://doi.org/10.5555/ajhb.2007.31.supp.S69 PMID: 17931139

132. Noar SM, Benac CN, Harris MS. Does tailoring matter? Meta-analytic review of tailored print health behavior changes interventions. Psychol Bull 2007; 133:673. <u>https://doi.org/10.1037/0033-2909.133.4</u>. 673 PMID: 17592961

133. Prasad PVS et al. MDT-MB therapy in paucibacillary leprosy: A clinicopathological assessment. Study. 71 (4); 2005: 242-245.

134. Reddy VM et al. Antituberculosis Activities of Clofazimine and Its New Analogs B4154 and B4157. Antimicrobagents Chemother. 40 (3); 1996: 633-636.

135. Makarov V et al. Synthesis and antileprosy activity of some dialkyldithiocarbamates. J Antimicrob Chemother . 57 (4); 2006:1134-1138.

136. Bacman D et al. Dapsone and Retinoids. 2001: 373-390.

137. Lucia M, Penna F. Considerations in the design of clinical trials for multibacillary leprosy treatment. 4 (1): 2014:77-86.

138. Ferreira DA et al. Analysis of the molecular association of rifampicin with hydroxypropyl- \$ -cyclodextrin. Brazilian J Pharm Sci. 40; 2004: 43-51.

139. WHO Model Prescribing Information, Drugs Used in Leprosy.1998: 1-32.

140. Subhashree Sahoo et al. Research Journal of Pharmaceutical Biological and Chemical Sciences Characterization of Controlled Release Ofloxacin Suspensions by Fourier. Res J Pharm Biol Chem Sei. 2 (4); 2011: 926-939.

141. F.J.Scholes G. Prednisolone Syrup USP. 1999

142. Mittal J, Sharma MM, Batra A. Tinospora cordifolia: amultipurpose medicinal plant- A. J Med Plants Stud. 2 (2); 2014:32-47.

143. Nishi Gupta et al. Medicinal Value of Euphorbia Tirucalli. Syst Rev Pharm. 4 (1); 2013: 40-46.

144. Thomas PPJJ, Skaria SMBP. Medicinal plants. 1998.

145. Mst Nazma Yesmin et al. Antioxidant and Antibacterial Activities of Calotropis procera Lin. Am-Euras J Agric Environ Sci. 4 (5); 2008: 550-553.

146. C tora et al. An overview on phytochemical and pharmacological profile of Cassia tora Linn. Int J Herb Med. 4(6): 2016: 50-55.

147. Azmi Let al. Pharmacological and biological overview on Mimosa pudica Linn. Int J Pharm LIFE Sei. 2 (11): 2011: 1226-1234.

148. Mustapha AA et al. Plant Remedies Practiced by Ketti People in the Management of Dermatosis. J Med Plants Stud Plant. 1 (5);2013: 112-118.

149. Sharma S, Kumar A. Tribal uses of medicinal plants of rajashthan : kachnar . Int J life Sci pharma Res. 2 (4); 2012: 70-76.

150. Hope G. A Literature Survey of Studies Performed by Master Students at Department de Médecine Traditionelle (DMT) inBamako, Mali. 2005.

151. Medicinal Plants of the area. 18-214.

152. Bharath M et al. Dalbergia sissoo DC. - An important medicinal plant. Int J Res Pharm Chem. 3 (2); 2013: 384-388.

153. Kumar KPS et al. Traditional and Medicinal Uses of Banana. J Pharmacogn Phytochem Traditz. 1 (3); 2012: 51-63. Extracts for In-vitro Antimicrobial Activity. Middle-East J Sci Res. 4 (4); 2009: 271-278.

155. Zhasa NN et al. Indigenous Knowledge on Utilization of plant Biodiversity for Treatment and Cure of diseases of Human beings in Nagaland, India: A case study. Int Res J Biol Se. 4 (4): 2015: 89-106.

156. Sahu PK et al. Ethnomedicinal Plants Used in the Healtheare Systems of Tribes of Dantewada. Am J Plant Sei. S; 2014: 1632-1643.

157. Koua FHM. Striga hermonthica (Del.) Benth: Phytochemistry and pharmacological properties outline. J Appl Pharm Sci. 1 (7): 2011: 1-5.

158. Ajibesin KK. Ethnobotanical survey of plants used for skin diseases and related ailments in. Ethnobot Res Appl.10; 2012: 463-522.

159. Okonogi S et al. Nanoencapsulation of Centella asiatica bioactive extract. In: XVIth International Conference on Bioencapsulation. 2008: 4-7.

160. Raman BV et al. Antibacterial Activities of Some Folk Medicinal Plants of Eastern Ghats. J Pure Appl Microbiol.3 (I); 2009: 187-194.

161. Varaprasad Bobbarala et al. Antifungal activity of selected plant extract against phytopathogenie fungi Aspergillus niger F2723. Indian J Sei Technol. 2 (4): 2009: 87-90.

162. Calapai G. Assessment Report on Centella Asiatica (L.) Urban, Herba. 2010: 1-44.

163. Prasad PVS et al. MDT-MB therapy in paucibacillary leprosy: A clinicopathological assessment. Study. 71 (4);2005: 242-245.

164. Reddy VM et al. Antituberculosis Activities of Clofazimine and Its New Analogs B4154 and B4157. Antimicrobagents Chemother. 40 (3); 1996: 633-636.

165. Makarov V et al. Synthesis and antileprosy activity of some dialkyldithiocarbamates. J Antimicrob Chemother.57 (4): 2006: 1134-1138.

166. Bacman D et al. Dapsone and Retinoids. 2001: 373-390.

167. Lucia M, Penna F. Considerations in the design of clinical trials for multibacillary leprosy treatment. 4 (1): 2014:77-86.

168. Ferreira DA et al. Analysis of the molecular association of rifampicin with hydroxypropyl- B -cyclodestrin. Brazilian J Pharm Sci. 40; 2004: 43-51.

169. WHO Model Prescribing Information, Drugs Used in Leprosy. 1998: 1-32.

170. Subhashree Sahoo et al. Research Journal of Pharmaceutical, Biological and Chemical Sciences Characterization of Controlled Release Ofloxacin Suspensions by Fourier. Res J Pharm Biol Chem Sci. 2 (4): 2011: 926-939.

171. F.J.Scholes G. Prednisolone Syrup USP. 1999.